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07/20/00**UTILITY PATENT APPLICATION TRANSMITTAL****(Large Entity)**

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Docket No.

RDID0050US

Total Pages in this Submission

31

**TO THE ASSISTANT COMMISSIONER FOR PATENTS**

Box Patent Application

Washington, D.C. 20231

Transmitted herewith for filing under 35 U.S.C. 111(a) and 37 C.F.R. 1.53(b) is a new utility patent application for an invention entitled:

**METHOD AND ARRANGEMENT FOR DETERMINING THE CONCENTRATION OF GLUCOSE IN A BODY FLUID**

and invented by:

**Udo HOSS, Hans-Joerg PFLEIDERER, Ralf GESSLER, Hans-Ulrich ZIETEN and Rolf FUSSGAENGER**

If a CONTINUATION APPLICATION, check appropriate box and supply the requisite information:

Continuation    Divisional    Continuation-in-part (CIP)   of prior application No.: \_\_\_\_\_

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Continuation    Divisional    Continuation-in-part (CIP)   of prior application No.: \_\_\_\_\_

which is a:

Continuation    Divisional    Continuation-in-part (CIP)   of prior application No.: \_\_\_\_\_

Enclosed are:

**Application Elements**

1.  Filing fee as calculated and transmitted as described below

2.  Specification having 20 pages and including the following:

- a.  Descriptive Title of the Invention
- b.  Cross References to Related Applications (*if applicable*)
- c.  Statement Regarding Federally-sponsored Research/Development (*if applicable*)
- d.  Reference to Microfiche Appendix (*if applicable*)
- e.  Background of the Invention
- f.  Brief Summary of the Invention
- g.  Brief Description of the Drawings (*if drawings filed*)
- h.  Detailed Description
- i.  Claim(s) as Classified Below
- j.  Abstract of the Disclosure

**UTILITY PATENT APPLICATION TRANSMITTAL****(Large Entity)**

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31

**Application Elements (Continued)**

3.  Drawing(s) (when necessary as prescribed by 35 USC 113)  
a.  Formal Number of Sheets 2  
b.  Informal Number of Sheets \_\_\_\_\_

4.  Oath or Declaration  
a.  Newly executed (*original or copy*)  Unexecuted  
b.  Copy from a prior application (37 CFR 1.63(d)) (*for continuation/divisional application only*)  
c.  With Power of Attorney  Without Power of Attorney  
d.  DELETION OF INVENTOR(S)  
Signed statement attached deleting inventor(s) named in the prior application,  
see 37 C.F.R. 1.63(d)(2) and 1.33(b).

5.  Incorporation By Reference (*usable if Box 4b is checked*)  
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under  
Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby  
incorporated by reference therein.

6.  Computer Program in Microfiche (*Appendix*)

7.  Nucleotide and/or Amino Acid Sequence Submission (*if applicable, all must be included*)  
a.  Paper Copy  
b.  Computer Readable Copy (*identical to computer copy*)  
c.  Statement Verifying Identical Paper and Computer Readable Copy

**Accompanying Application Parts**

8.  Assignment Papers (*cover sheet & document(s)*)

9.  37 CFR 3.73(B) Statement (*when there is an assignee*)

10.  English Translation Document (*if applicable*)

11.  Information Disclosure Statement/PTO-1449  Copies of IDS Citations

12.  Preliminary Amendment (*to follow*)

13.  Acknowledgment postcard

14.  Certificate of Mailing

First Class  Express Mail (*Specify Label No.:* EL566320556US)

**UTILITY PATENT APPLICATION TRANSMITTAL**  
**(Large Entity)**

(Only for new nonprovisional applications under 37 CFR 1.53(b))

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**RDID0050US**

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**31**

**Accompanying Application Parts (Continued)**

15.  Certified Copy of Priority Document(s) (*if foreign priority is claimed*)

16.  Additional Enclosures (*please identify below*):

**General Appointment of Representative for U.S. Patent and Patent Application.**

**Fee Calculation and Transmittal**

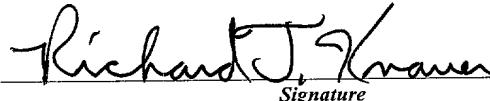
**CLAIMS AS FILED**

For	#Filed	#Allowed	#Extra	Rate	Fee
<b>Total Claims</b>		- 20 =	0	x \$18.00	\$0.00
<b>Indep. Claims</b>		- 3 =	0	x \$78.00	\$0.00
<b>Multiple Dependent Claims (check if applicable)</b>					\$0.00
				<b>BASIC FEE</b>	\$0.00
<b>OTHER FEE (specify purpose)</b>					\$0.00
				<b>TOTAL FILING FEE</b>	\$0.00

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*Richard T. Knauer*  
 Signature

Richard T. Knauer, Reg. No. 35,575  
 Roche Diagnostics Corporation  
 9115 Hague Road, Bldg. D  
 P.O. Box 50457  
 Indianapolis, IN 46250-0457  
 Telephone No.: (317) 576-7464  
 Facsimile No.: (317) 576-2883

Dated: July 20, 2000

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**CERTIFICATE OF MAILING BY "EXPRESS MAIL" (37 CFR 1.10)**

Applicant(s): Udo HOSS, et al.

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To Be AssignedFiling Date  
July 20, 2000Examiner  
To Be AssignedGroup Art Unit  
To Be AssignedInvention: **METHOD AND ARRANGEMENT FOR DETERMINING THE CONCENTRATION OF GLUCOSE IN A BODY FLUID**

I hereby certify that the following correspondence:

**Utility Patent Application***(Identify type of correspondence)*is being deposited with the United States Postal Service "Express Mail Post Office to Addressee" service under  
37 CFR 1.10 in an envelope addressed to: The Assistant Commissioner for Patents, Washington, D.C. 20231 onJuly 20, 2000.*(Date)***Rose Edwards***(Typed or Printed Name of Person Mailing Correspondence)**(Signature of Person Mailing Correspondence)***EL566320556US***("Express Mail" Mailing Label Number)*

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**Method and arrangement for determining the concentration of glucose in a body fluid**

**Description**

The invention concerns a method and an arrangement for determining the concentration of glucose in a body fluid, in particular in tissue fluid according to the preamble of the patent claims 1 or 14 or 22.

A process and an arrangement of this type are known from WO 97/42868. This proposes intermittent delivery pulses in order to, on the one hand, enable a continuous signal calibration and, on the other hand, to accelerate the measurement process. In this process the perfusate volume which is at that moment in the microdialysis probe adjusts during the resting phases between the delivery pulses to the concentration of the tissue glucose as a result of the dialysis process whereas adjacent volume regions in the subsequent liquid column that is transported further at a high flow rate remain largely unchanged. A signal peak is observed during a delivery pulse in the measuring cell which corresponds to the concentration gradient and from which the glucose content of the dialysate and thus also of the body fluid can be determined. Perfusion fluid containing glucose, the specified glucose concentration of which defines the baseline value of the signal peak, is used for the calibration. A prerequisite for this is a linear sensor behaviour during the dialysis phases in addition to a complete concentration equilibration and it is assumed that the concentration profile in the volume transported away from the probe does not decay until it reaches the measuring cell. However, especially the latter

assumption is frequently not the case since mixing occurs especially when the flow is laminar. In addition diffusion exchange disturbs the glucose equilibrium in the tissue surrounding the probe.

Taking this as a starting point the object of the invention is to avoid the aforementioned disadvantages and in particular concentration disturbances in the body fluid and to enable an exact glucose determination with a reduced dialysis period.

The feature combinations stated in the claims 1, 14 and 22 are proposed to achieve this object. Advantageous embodiments and further developments of the invention derive from the dependent claims.

The idea behind the invention is to adapt the glucose content of the perfusate in a self-adjusting and adaptive manner to the glucose concentration of the body fluid. Accordingly the method proposed to achieve the above-mentioned object is to adapt the initial content of the glucose in the perfusate to the glucose content of the body fluid by means of a control unit according to a command variable derived from the measurement signals of the measuring cell. This offsets glucose gradients and hence reduces the period required for a complete dialysis equilibration. It also avoids interfering effects due to glucose gradients even with a high flow rate through the microdialysis probe and glucose variations in the body fluid.

A particularly preferred embodiment of the invention envisages determination of the momentary starting content of glucose in the perfusate as a measure for the glucose content of the body fluid when the deviation is

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negligible. This enables a quantitative indirect determination of the concentration by means of the momentary actual value of the regulating variable while the continuously measured signals from the measuring cell are only used as regulating input variables. Alternatively additionally, it is in principle possible to derive the glucose content of the body fluid directly from the measurement signals.

The initial content of glucose in the perfusate is advantageously determined from the adjustable variable of the adjuster of the controlling device. This measure enables the initial content to be determined accurately for example by comparison with normalized values in tables without requiring additional glucose sensors. However, in principle it is also possible to measure the glucose content of the perfusate before it is passed into the microdialysis probe.

For a variable adjustment it is advantageous when the initial content of glucose in the perfusate is influenced by flow mixing two perfusion fluids with different glucose concentrations that are kept ready in two separate reservoirs.

In a particularly preferred embodiment of the invention the perfusate is passed through the microdialysis probe in alternating consecutive transport and dialysis intervals at different flow rates, the flow rate during the transport intervals being higher than during the dialysis intervals. As a result the measurement can be shortened as a whole and the evaluation can be further simplified since an existing concentration gradient can be qualitatively detected by means of the measurement signal even if there is only a partial dialysis

equilibration. The flow rate should be increased during the transport intervals to such an extent that the starting content of the glucose in the perfusate is essentially maintained as it passes through the microdialysis probe. In contrast the transport is interrupted or at least the flow rate is reduced during the dialysis intervals to such an extent that the glucose concentration in the dialysate approximates the glucose content of the body fluid.

In a particularly simple control process the command variable which defines the target value is determined by integration or differentiation of the time course of the measurement signals or by a qualitative detection of signal peaks in the time course of the measurement signals. Alternatively the command variable can be determined by comparing the actual signal time course of the measurement signals with calibrated signal patterns deposited in a storage medium. An additional method is to determine the command variable from the peak value of the signal time course of the measurement signals during each transport interval. In order to quantitatively define the regulating input signal, the command variable can be determined according to the glucose content  $c$  of the body fluid using the relationship

$$c = \left[ \frac{S_g}{S_g \cdot (1-b) + b \cdot S_0} - 1 \right] \cdot a \cdot c_0 + c_0$$

in which  $S_g$  denotes the peak value and  $S_0$  denotes the base line value of the signals measured during a transport interval and  $c_0$  is the momentary starting content of glucose in the perfusate and  $a$ ,  $b$  are empirically determined correction factors compensating for diffusion and mixing and remaining recovery effects during the transport interval.

A particularly simple control function envisages that the initial content of glucose in the perfusate is adjusted discontinuously by a two-point control process in which the starting content of the glucose in the perfusate is changed by a predetermined adjusting value when there is a deviation.

With regard to a measuring arrangement, a control device is proposed to achieve the above-mentioned object which adapts the starting content of the glucose in the perfusate to the glucose content of the body fluid on the basis of a command variable derived from the measurement signals of the measuring cell. An evaluation unit is provided in a preferred embodiment which determines the glucose content of the body fluid corresponding to the momentary starting content of glucose in the perfusate when the deviation is negligible.

The perfusion device contains a store of perfusate and a transport unit to transport perfusate. The transport unit preferably operates at intervals i.e. at different delivery rates in successive time intervals. In order to vary the initial glucose content it is advantageous if the perfusate store has at least two separate reservoirs to hold perfusion liquids with different glucose concentrations. Advantageously the perfusate store has a first reservoir containing a glucose-free perfusion liquid and a second reservoir containing a glucose-containing perfusion liquid. In this case the glucose content in the latter should be above the physiological thresholds. A controller for adjusting the starting content of the glucose in the perfusate which is simple to construct is preferably provided by a flow mixer composed of a mixing valve or a clock-pulsed directional

control valve as the adjuster. In this case it is advantageous for the flow mixer to be connected on the inlet side with at least two reservoirs to supply perfusion fluids with different glucose contents and to discharge into a perfusate tube leading to the microdialysis probe.

The control device advantageously has a controller that operates digitally preferably by means of a microcontroller in order to process the signal flow in a variable manner.

The invention is elucidated in more detail in the following on the basis of an embodiment example shown schematically in the drawing.

Fig. 1 shows a block diagram of a microdialysis arrangement for determining the concentration of glucose in a tissue fluid and

Fig. 2 shows a time-dependency diagram of the perfusate flow, the measured glucose signal in the dialysate and the adaptively readjusted glucose concentration in the perfusate.

The microdialysis arrangement shown in the figure is essentially composed of a microdialysis probe 10 that can be implanted in the subcutaneous tissue of a test person, a perfusion device 12, 14 for the intermittent perfusion of the microdialysis probe 10 with glucose-containing perfusate, a flow-through measuring cell 16 to detect the glucose content in the dialysate that flows through, a control device 18, 20 to adjust the starting content of the glucose in the perfusate to the

glucose content of the tissue fluid and an evaluation unit 22 to determine the glucose content of the tissue fluid.

The microdialysis probe 10 has a dialysis membrane 24 which enables a diffusion exchange of glucose between the perfusate located in the probe and the interstitial liquid surrounding the probe while obtaining dialysate. For this purpose a flow-through channel is provided in the tube-shaped double bore probe housing 25 which is at least partially bordered by the dialysis membrane 24 and which in the proximal probe region is connected on the inlet side with a perfusate tube 26 to pass in perfusate and on the outlet side with a dialysate tube 28 to discharge the dialysate formed from the perfusate during the dialysis process. The dialysate can be further transported via the dialysate tube 28 to the measuring cell 16 and from there into a collecting vessel 30. Suitable microdialysis probes of the described type are well-known especially from DE-A 33 42 170 or US-PS 4,694,832 and can be obtained from the CMA/Microdialysis AB company located in Solna, Sweden under the name "CMA 60 Microdialysis Catheter" or "CMA 70 Brain Microdialysis Catheter".

In order to supply the microdialysis probe 10 at intervals with perfusate containing glucose, the perfusion device contains a store of perfusate 12 and a transport unit 14. The perfusate store 12 is composed of two separate reservoirs 32, 34 one of which contains a glucose-free perfusion liquid 36 and the other contains a perfusion liquid 38 to which glucose has been added at a specified concentration. The glucose concentration of the liquid 38 is expediently more than 4 g/l in order that the physiological range of tissue glucose can be

covered in the perfusate by mixing the liquids 36 and 38 in the manner described below. A peristaltic pump 14 operated at intervals is provided as a feeding unit to transport the perfusate in metered delivery pulses of a few microlitres through the microdialysis probe 10 and the subsequent measuring cell 16. It is preferably located in the dialysate tube 28 in order during the transport pauses to isolate the microdialysis probe 10 from the measuring cell 16 arranged extracorporeally.

The measuring cell 16 through which perfusion fluid and the dialysate contained therein flows, has an electrode sensor 40 which operates electrochemically-enzymatically for continuous signal acquisition. The sensor 40 has a measuring electrode that is not shown which is supplied with the dialysate that serves as the electrolyte and is used to continuously register as a continuous measuring current, the measurement signals that are linearly dependent on the glucose content of the dialysate. Further details of this measuring principle are known in the prior art in particular from DE-A 44 01 400 to which special reference is herewith made. Obviously the measurement signals also reflect the glucose content of the body fluid insofar as a complete equilibration of the concentration gradient between the perfusate and the body fluid has taken place in the microdialysis probe or the degree of equilibration is known.

The signals measured by the sensor 40 are electronically processed in the subsequent measurement transducer 42 and fed into a digital controller 18 of the controlling device as a chronological sequence of digital values by means of a clock-pulsed analogue-digital converter. In this case the controller 18 is a microcontroller which also forms the evaluation unit 22. The output side of

the controller 18 is connected to a directional control valve 20 as an adjuster of the control unit for adjusting the initial content of glucose in the perfusate. In a spring-centred first switch position the directional control valve 20 connects the perfusate tube 26 to the glucose-free reservoir 32 and, in a second electromagnetically actuated switch position, to the glucose-containing reservoir 34. Hence, the glucose concentration in the perfusate can be influenced as a regulated variable in a synchronized operation by suitable selection of the switching frequency, by the quantity ratio of the liquids 36, 38 sucked in by the peristaltic pump 14 and by the flow mixing which occurs subsequently in the perfusate tube 26.

When the microdialysis arrangement is in operation according to the above-mentioned diagram in fig. 2, the perfusate is pumped in transport intervals 46 separated by resting and dialysis intervals 44 through the microdialysis probe 10 and the measuring cell 16. The dialysis intervals can be dimensioned such that the glucose content of the perfusate volume resting in the microdialysis probe 10 is almost completely adjusted to the tissue glucose by diffusion exchange. In contrast the glucose concentration in the perfusate remains essentially unchanged during the transport intervals 46 due to rapid passage through the probe. The degree of equilibration or the recovery depends among others on the residence time and the flow rate of the perfusate in the microdialysis probe 10. In the embodiment example shown in fig. 2 the duration of the dialysis interval is 360 sec when transport is interrupted whereas the duration of a transport interval is 180 sec at a flow rate  $\dot{V}$  of 0.08  $\mu\text{l}/\text{s}$ .

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With each delivery pulse the dialysate formed in the previous dialysis interval is displaced completely in a transport or liquid column from the microdialysis probe 10 at least into the dialysis tube 28 and preferably up to the measuring cell 16. Accordingly a signal S is registered there during the transport intervals 46 which, when there are differences in concentration between the tissue and perfusate glucose, displays a corresponding peak or extreme value  $S_g$  and a base line value  $S_0$  which corresponds to the starting concentration of glucose in the perfusate (middle diagram in fig. 2).

In order to adjust the starting glucose in the perfusate to the tissue glucose, a command variable which corresponds to the tissue glucose is derived from the measurement signals by means of the evaluation unit 22 and fed into the controller 18 to generate the deviation from the controlled variable i.e. the momentary value for the starting glucose. In this case the command variable correlates with the signal peaks  $S_g$  whereas the control variable can be acquired from the base line value  $S_0$ .

For a particularly simple control it is sufficient that the command variable or the control difference is determined by a qualitative detection of signal peaks  $S_g$  and the starting content of glucose in the perfusate is discontinuously adapted by a two-point control process. In this process the initial glucose content  $c_p$  is increased by a predetermined value  $\Delta p$  when there is a positive peak (signal peak 48) using the adjuster 20 and is correspondingly reduced when there is a negative peak (signal dip; not shown). A slight equilibration or recovery (< 50 %) during the dialysis intervals is

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already sufficient for this controlled operation and hence its duration can be correspondingly reduced.

The adjusting signal can only be converted in the case of constant transport intervals after a dialysis interval is completed. In order to avoid this dead time, it is also conceivable that the duration of the momentary transport interval could be extended when there is a deviation to ensure that the perfusion liquid containing the re-adjusted glucose content can immediately pass into the microdialysis probe 10.

When the deviation is negligible, a constant signal 50 is finally observed which indicates that the initial glucose content  $c_p$  agrees with the actual value  $c_g$  of the tissue glucose (fig. 3 below). Hence an error-prone direct evaluation of the measurement signals is not necessary and the method enables the tissue glucose to be determined indirectly from the equilibrium values  $c_p$  when a constant signal 50 occurs. This can be accomplished without additional measurements by the fact that the initial content of the glucose in the perfusate is determined by means of the evaluation unit 22 from the actual adjustable variable i.e. the switching frequency of the valve 20, optionally by comparison with assigned calibration values.

One method for the quantitative evaluation of the measurement signals is to determine the command variable by pattern recognition i.e. by comparing the actual signal curve of the measurement signals with calibrated signal patterns that are deposited in a storage medium. Alternatively the deviation can be determined as a difference between the peak value and the base line value of the signal curve of the measurement signals.

Hence in this case the actual values of the control variable are recorded by measurement as the base line value  $S_0$ .

As outlined above, the sensor signal is evaluated only during the high flow phase. This signal consists of the part  $S_g$  belonging to the dialysis phase and being proportional to the tissue glucose concentration and the signal  $S_0$  belonging to the high flow rate representing almost the initial perfusate glucose concentration. In order to achieve a control which is independent of variations in sensitivity, the command variable can be determined according to the glucose content  $c$  of the tissue liquid using the relationship

$$c = \frac{S_g}{S_0} * c_0$$

in which  $c_0$  is the momentary starting glucose content in the perfusate which can be determined by the adjustable variable.

In order to take into account further influence parameters, the tissue glucose concentration may be calculated as follows:

$$c = \left[ \frac{S_g}{S_g \cdot (1-b) + b \cdot S_0} - 1 \right] \cdot a \cdot c_0 + c_0$$

where  $a$  and  $b$  are empirically determined correction factors compensating for diffusion and mixing and remaining recovery effects during the transport interval. The peak value  $S_g$  is the sensor value at a definite time point during the transport interval. This time point may be determined *in vitro* and is given by the time it takes to push the dialysate from the microdialysis probe to the glucose sensing unit, i.e. the measuring cell 16.  $S_0$  may be obtained as the mean

value of the sensor values 60 seconds before and after the time point of  $S_g$ .

It is conceivable that the previously described principle of self-adapting control of the glucose content can also be used in the case of measurements with a continuous perfusate flow. In principle this microdialysis technique is not restricted to subcutaneous measurements on the human body. Rather it is possible to examine other body fluids such as blood and optionally ex vivo.

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**Claims**

What is claimed is:

1. Method for determining the glucose concentration in a body fluid, in particular tissue fluid, in which perfusate containing glucose is passed through a microdialysis probe (10) inserted into the body fluid and dialysate obtained in this process is transported to a measuring cell (16), and in which measurement signals that correlate with the glucose content of the dialysate are measured at the measuring cell (16), **characterized in that** the starting content of glucose in the perfusate is adjusted to the glucose content of the body fluid by means of a control device (18, 20) in accordance with a command variable derived from the measurement signals of the measuring cell (16).
2. Method as claimed in claim 1, **characterized in that** when the control deviation is negligible the momentary starting content of the glucose in the perfusate is determined as a measure for the glucose content of the body fluid.
3. Method as claimed in claims 1 or 2, **characterized in that** the starting content of glucose in the perfusate is determined from the adjusting variable of an adjuster (20) of the control device (18, 20).

4. Method as claimed in one of the claims 1 to 3, **characterized in that** the glucose content of the perfusate is measured before it is passed into the microdialysis probe (10).

5. Method as claimed in one of the claims 1 to 4, **characterized in that** the starting content of glucose in the perfusate is influenced by flow mixing two perfusion liquids (36, 38) with different glucose concentrations provided in two separate reservoirs (32, 34).

6. Method as claimed in one of the claims 1 to 5, **characterized in that** the perfusate is passed through the microdialysis probe (10) in alternating successive transport and dialysis intervals at different flow rates, the flow rate during the transport intervals being higher than during the dialysis intervals.

7. Method as claimed in claim 6, **characterized in that** the flow rate during the transport intervals is increased to such an extent that the starting content of glucose in the perfusate during passage through the microdialysis probe (10) remains essentially constant and that during the dialysis intervals the transport is interrupted or at least the flow rate is reduced to such an extent that the glucose concentration of the dialysate approximates the glucose content of the body fluid.

8. Method as claimed in one of the claims 1 to 7, **characterized in that** the command variable is determined by integration or differentiation of the time course of the measurement signals.

9. Method as claimed in one of the claims 6 to 8,  
**characterized in that** the command variable is  
determined by qualitative detection of signal peaks  
in the time course of the measurement signals.

10. Method as claimed in one of the claims 6 to 9,  
**characterized in that** the command variable is  
determined by comparing the actual signal curve of  
the measurement signals with calibrated signal  
patterns deposited in a storage medium.

11. Method as claimed in one of the claims 6 to 10,  
**characterized in that** the command variable is  
determined from the peak value of the signal time  
course of the measurement signals during each  
transport interval.

12. Method as claimed in one of the claims 6 to 11,  
**characterized in that** the command variable can be  
determined according to the glucose content  $c$  of  
the body fluid according to the relationship

$$c = \left[ \frac{S_g}{S_g \cdot (1-b) + b \cdot S_0} - 1 \right] \cdot a \cdot c_0 + c_0$$

in which  $S_g$  denotes the peak value and  $S_0$  denotes  
the base line value of the signals measured during  
a transport interval and  $c_0$  is the momentary  
starting content of glucose in the perfusate and  $a$ ,  
 $b$  are empirically determined correction factors  
compensating for diffusion and mixing and remaining  
recovery effects during the transport interval.

13. Method as claimed in one of the claims 1 to 12, **characterized in that** the starting content of glucose in the perfusate is regulated discontinuously by a two-point control process in which the starting content of glucose in the perfusate is changed by a predetermined adjustment value when there is a control deviation.

14. Arrangement for determining the glucose concentration in a body fluid, in particular tissue fluid, comprising a microdialysis probe (10) for the diffusion exchange of glucose with surrounding body fluid, a perfusion device (12, 14) for perfusing the microdialysis probe (10) with glucose-containing perfusate to obtain dialysate and a measuring cell (16) located after the microdialysis probe (10) for detecting measurement signals that correlate with the glucose content of the dialysate, **characterized by** a control device (18, 20) which adjusts the starting content of glucose in the perfusate to the glucose content of the body fluid in accordance with a command variable derived from the measurement signals of the measuring cell (16).

15. Arrangement as claimed in claim 14, **characterized by** an evaluation unit (22) to determine the momentary starting content of glucose in the perfusate when the control deviation is negligible as a measure for the glucose content of the body fluid.

16. Arrangement as claimed in claims 14 or 15, **characterized in that** the perfusate device has a perfusate store (12) and a transport unit (14) for the preferably intermittent transport of perfusate.

17. Arrangement as claimed in claim 16, **characterized in that** the perfusate store (12) has at least two separate reservoirs (32, 34) to hold perfusion liquids (36, 38) with different glucose concentrations.

18. Arrangement as claimed in claim 16 or 17, **characterized in that** the perfusate store (12) has a first reservoir (32) containing a glucose-free perfusion liquid (36) and a second reservoir (34) containing a glucose-containing perfusion liquid (38).

19. Arrangement as claimed in one of the claims 14 to 18, **characterized in that** the control device has a flow mixer (20) preferably comprising a mixing valve or clock-pulsed directional control valve as an adjuster to adjust the starting content of glucose in the perfusate.

20. Arrangement as claimed in claim 19, **characterized in that** the inlet side of the flow mixer (20) is connected to at least two reservoirs (32, 34) for feeding in perfusion fluids with different glucose contents and the outlet side of the flow mixer (20) connects a perfusate tube (26) leading to the microdialysis probe (10).

21. Arrangement as claimed in one of the claims 14 to 20, **characterized in that** the control device has a digitally operated controller (18) preferably in the form of a microcontroller.

22. Arrangement for determining the glucose concentration in a body fluid, in particular tissue fluid, comprising a microdialysis probe (10) inserted into the body fluid, at least two reservoirs (32, 34) for holding perfusion liquids (36, 38) with different glucose contents, a transport unit (14) to perfuse the microdialysis probe (10) with glucose-containing perfusate to obtain dialysate and a flow-through measuring cell (16) located downstream of the microdialysis probe (10) to register measurement signals that correlate with the glucose content of the dialysate, **characterized by** a control device (18, 20) connected on the input side to the measuring cell (16) which control device has a flow mixer (20) connected on the inlet side to the reservoirs (32, 34) and on the outlet side to the microdialysis probe (10) which acts as an adjuster to regulate the starting content of glucose in the perfusate.

## **Abstract**

### Method and arrangement for determining the concentration of glucose in a body fluid

The invention concerns a method and an arrangement for determining the concentration of glucose in a body fluid. In the microdialysis technology used for this purpose, perfusate containing glucose is transported in intermittent delivery pulses through a microdialysis probe (10) inserted into the body fluid and dialysate obtained in this process is passed to a measuring cell (16) to record the glucose content. In order to achieve an exact determination of glucose even with a reduced dialysis period, it is proposed that the starting content of glucose in the perfusate is adapted to the glucose content of the body fluid by means of a control device (18, 20) in accordance with a command variable derived from the measurement signals of the measuring cell (16). When the control deviation is negligible the momentary starting content of glucose in the perfusate can be determined as a measure for the glucose content of the body fluid.

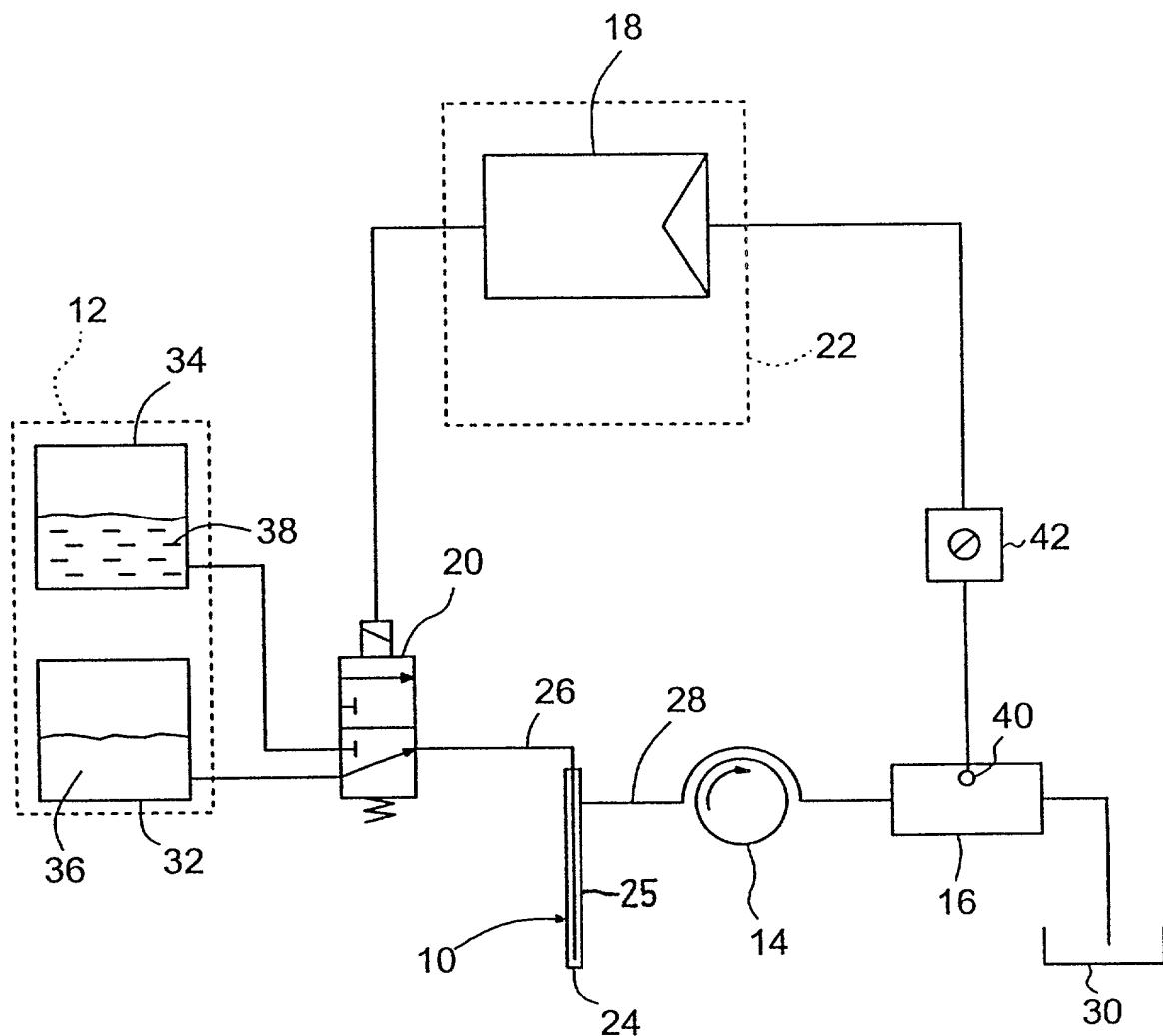


Fig. 1

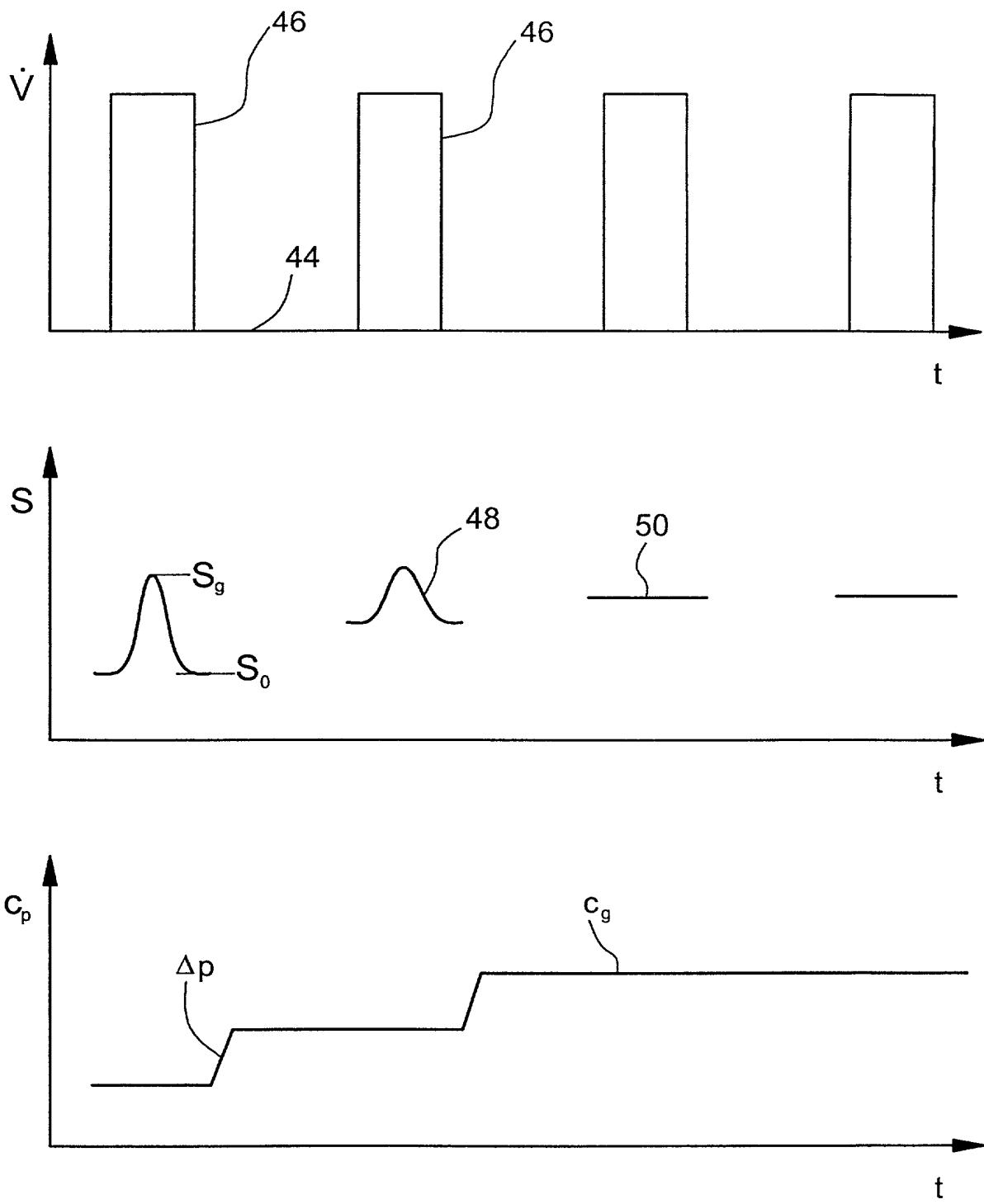


Fig. 2



I hereby claim the benefit under 35 U.S.C. Section 119(e) of any United States provisional application(s) listed below:

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

(Application Serial No.)

(Filing Date)

I hereby claim the benefit under 35 U. S. C. Section 120 of any United States application(s), or Section 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of 35 U.S.C. Section 112, I acknowledge the duty to disclose to the United States Patent and Trademark Office all information known to me to be material to patentability as defined in Title 37, C. F. R., Section 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application:

(Application Serial No.)

(Filing Date)

(Status)

(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)

(patented, pending, abandoned)

(Application Serial No.)

(Filing Date)

(Status)

(patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

**POWER OF ATTORNEY:** As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith. (*list name and registration number*)

**Richard T. Knauer, Reg. No. 35,575**

**D. Michael Young, Reg. No. 33,819**

**Brent A. Harris, Reg. No. 39,215**

**Kenneth J. Waite, Reg. No. 45,189**

**Marilyn L. Amick, Reg. No. 30,444**

Send Correspondence to: **Richard T. Knauer**  
**Roche Diagnostics Corporation**  
**9115 Hague Road, Bldg. D, P.O. Box 50457**  
**Indianapolis, IN 46250-0457**

Direct Telephone Calls to: (*name and telephone number*)

**Richard T. Knauer, Telephone No. (317) 576-7464**

Full name of sole or first inventor <b>HOSS, Udo</b>	Date
Sole or first inventor's signature	Date
Residence <b>24442 Valencia Blvd. #2102, Valencia, CA 91355, U.S.A.</b>	
Citizenship <b>German</b>	
Post Office Address <b>(same as above)</b>	

Full name of second inventor, if any <b>PFLEIDERER, Hans-Joerg</b>	Date
Second inventor's signature	Date
Residence <b>Lange Lemppen 5, D-89075 Ulm, Germany</b>	
Citizenship <b>German</b>	
Post Office Address <b>(same as above)</b>	

Full name of third inventor, if any <b>GESSLER, Ralf</b>	Date
Third inventor's signature	
Residence <b>Birkenstrasse 9, D-88255 Baienfurt, Germany</b>	
Citizenship <b>German</b>	
Post Office Address <b>(same as above)</b>	

Full name of fourth inventor, if any <b>ZIETEN, Hans-Ulrich</b>	Date
Fourth inventor's signature	
Residence <b>Vorderer Berg 34, D-89186 Illerrieden, Germany</b>	
Citizenship <b>German</b>	
Post Office Address <b>(same as above)</b>	

Full name of fifth inventor, if any <b>FUSSGAENGER, Rolf</b>	Date
Fifth inventor's signature	
Residence <b>Siedlerweg 7, D-89134 Blaustein-Weidach, Germany</b>	
Citizenship <b>German</b>	
Post Office Address <b>(same as above)</b>	

Full name of sixth inventor, if any	
Sixth inventor's signature	Date
Residence	
Citizenship	
Post Office Address	

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Assistant Commissioner for Patents  
Washington, DC 20231

**GENERAL APPOINTMENT OF REPRESENTATIVE FOR**  
**U.S. PATENT AND PATENT APPLICATION**

The undersigned applicant or assignee hereby appoints D. Michael Young, Reg. No. 33,819, Richard T. Knauer, Reg. No. 35,575, Brent A. Harris, Reg. No. 39,215, Jill Lynn Woodburn, Reg. No. 39,874, and Marilyn L. Amick, Reg. No. 30,444 all of Roche Diagnostics Corporation, 9115 Hague Road, P.O. Box 50457, Indianapolis, Indiana 46250, Telephone (317) 845-2000:

to prosecute and transact all business on its behalf before the United States Patent and Trademark Office in connection with any U.S. patent assigned to it and any U.S. patent application filed by it or on its behalf and to receive payments on its behalf.

Signed this 24th day of November, 1999 at Mannheim, Germany.

Roche Diagnostics GmbH



Signature

Dr. Bernd Kolb

Print Name

Senior Vice President

Position or Title

Roche Diagnostics GmbH



Signature

Dr. Andreas Poreda

Print Name

Director

Position or Title